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Biochar and Biochar Magnetite of *Spirulina platensis* to Remove Cadmium Ions from Aqueous Solutions

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ABSTRACT

An investigation was conducted using both biochar and magnetic biochar. Have been utilized as an effective, affordable, secure, and ecologically friendly way to get rid of heavy metals like cadmium. Microalgae from Spirulina platensis were used to make biochar by burning it. We mix biochar with hydrogen peroxide to create functional groups on the surface by using ice bath sonication which is biochar activate (BC). Also, magnetic biochar (MBC) has been prepared by mixing biochar activated with Fe₃O₄ (2:1 mole ratio) respectively. Two concentrations of aqueous solutions (5 ppm and 10 ppm) were mixed with two weights of biochar and magnetic biochar (0.1 and 0.5 g) for 10, 20, and 40 minutes. The concentration of cadmium (5 ppm) with 0.5 g of biochar (BC) after 40 minutes removed 97.58% of cadmium ion from the aqueous solution. On the other hand, magnetic biochar (MBC) (0.5 g) with 5 ppm of cadmium solution after 40 minutes could remove 100% of cadmium ions. We character BC, MBC, and biochar treated with cadmium ions by using FTIR, FESEM, EDX, and XRD studies. These analytical results indicate that the effective groups in biochar and magnetic biochar can adsorb heavy metals by carboxyl, hydroxyl, and carbonyl groups. The findings of the investigation on magnetic biochar revealed that, in addition to carbonyl, hydroxyl, and carboxylic groups, the substance also included magnetite. The findings made it very clear that removing cadmium with biochar reduced their risk and did it in a quick, inexpensive, and environmentally beneficial manner.

INTRODUCTION

Heavy metal contamination in drinking water and wastewater poses a major threat to the environment and all living things on land, in the air, and in the water. Heavy metals were processed using a variety of cutting-edge biological and conventional techniques based on nanomaterials. Microalgae are a significant class of microorganisms used in biological processes that can remove heavy metals from wastewater and have numerous environmental uses (Priatni *et al.*, 2018). One of the potential biological treatment methods is the use of microalgae to remove heavy metals from contaminated soil or water (Goswami *et al.*,2022). Polysaccharides, proteins, and lipids found on the surface of microalgae have a variety of functional groups, such as carboxyl, hydroxyl, carbonyl, and phosphodiester groups, that help heavy metal ions bind well (Pragya *et al.*, 2013). Algae are photosynthetic creatures that may live in freshwater, seawater, or sewage. In recent years, microalgae have been used to extract dangerous metals from wastewater treatment plants and transform them into less toxic molecules (Goh *et al.*,2022). Blue-green Spirulina platensis is a multicellular prokaryote (alga).

In order to bind heavy metals, its cell wall mostly consists of carbohydrates (cellulose and alginate) and organic lipids, which include a variety of functional groups (such as amino and carboxyl, hydroxyl, imidazole, phosphate, sulfonate, thiol, etc.) (Priatni et al., 2018). The stable carbon content of biochar is high, and it also has several nutrients. Because of its architecture with numerous micropores and resulting large surface areas, as well as its numerous functional groups, it is widely employed in carbon sequestration and soil improvement. Heavy metals in the soil and water can be removed using biochar as an adsorbent and residue (Qiu et al., 2022). The present study aims to determine the role of microalgae in the biological treatment of heavy metals, the mechanism of absorption of heavy metals, and the factors affecting the efficiency of removal.

MATERIALS AND METHODS

Spirulina platensis came from the

Star West Company in China. It was sieved and processed to make biochar. Other chemicals were purchased from Sigma-Aldrich (M) Sdn Bhd, including ethanol, sodium hydroxide, hydrochloric acid, cadmium sulfate, ferrous sulfate, and ferric chloride. The chemicals were used directly after receipt without any additional processing.

The two procedures that made up the assembly process for the composite magnetic biochar-based adsorbent (Scheme 1) were the production of MBC and biochar. Raw Spirulina platensis was first pyrolysis to create biochar utilizing a number of different input factors, including the pyrolysis temperature, heating time, and impregnation ratio (raw Spirulina platensis/H₂O₂). The obtained biochar was then combined with magnet materials to generate composite magnetic biochar. The biochar and magnetic biochar were used for the Cd2+ adsorption process after being analyzed.



Scheme 1: A summary diagram showing the preparation and testing of the composite magnetic biochar.

Biochar Preparation:

The Spirulina platensis was continuously cleaned with distilled water to

remove pollutants and dust from its surface and then dried for 24 hours at 105°C. The mesh size was decreased to the required 0.5 mm by passing the dried material through the proper sieves (Vibratory Sieve Shaker AS 200 Digit cA Model). It was then stored in an airtight container at room temperature until it was needed. Spirulina platensis was utilized to make charcoal by slow pyrolysis. We updated the preparation parameters in accordance with those that had been preoptimized in a tube furnace at a pyrolysis temperature of 600 °C, a reaction time of 4 hours (Kim *et al.*, 2019)..

Biochar Activation:

The next step is to create function groups on the surface of biochar by using hydrogen peroxide which is an alternative to the sodium hydroxide method (Kim et al., 2019). To prepare biochar to activate (BC activate), we disperse 1g of biochar in 20 mL of hydrogen peroxide (H₂O₂, 35%) then use bath sonication as the source of energy to create the hydroxyl, carboxyl and carbonyl temperature during groups. The the experiment must be between 10-25°C for 1h after that filtration, and the biochar activated on filter paper overnight to dry and washed with acetone three times.

Magnetic Preparation:

Magnetic iron oxide was created using the co-precipitation approach (Al-Kazazz et al., 2016). Fe₃O₄ MNPs are as follows: magnetic biochar was made using Fe₃O₄ nanoparticles. The magnetic nanoparticles were precipitated using methods that used ferrous sulfate (FeSO₄) and ferric chloride (FeCl₃.6H₂O) in a 2:1 ratio.

Preparation of Modified (magnetic) Biochar:

Following the production of MBC, biochar was added to 750 mL of deionized water by adding 5 g of Fe₃O₄ and 10 g of BC over a period of two hours. The magnetic biochar is dried for 24 hours at a temperature of 45 °C after being repeatedly washed with ethanol and then distilled water (Baltrenaite *et al.*, 2017).

Adsorption Study:

A 1000 ppm Cd^{2+} stock solution was prepared by dissolving 1.8 g of cadmium sulfate in deionized water. The stock solution was then diluted to different concentrations (5 and 10 ppm) for batch studies. The solution underwent a number of modifications to obtain the best adsorption performance. Batch adsorption studies were performed by adding biochar to a cadmium-aqueous solution in an Erlenmeyer flask. The adsorbent was used in doses of 0.1 and 0.5 g for a total of 10, 20, and 40 minutes.

$$R\% = \frac{(ci-cf)}{ci} \times 100 \dots (1)$$

Sorption capacity of biochar was calculated

according to the following formula:

$$A = \frac{c_1 - c_f}{m} \times v \ (mg. g^{-1})....(2)$$

Statically Analysis:

All data were analyzed using a univariate ANOVA. The probability threshold of 5% (P 0.05) was selected to illustrate the statistical difference. SPSS 26.0 (IBM, Chicago, IL, USA) is being used.

RESULTS AND DISCUSSION

At a concentration of 5 ppm, the highest rate of cadmium removal from water treated with biochar was found at a dose of (0.5) gm of biochar for (40) minutes. The lowest rate of cadmium removal was found at a dose of (0.1) gm of biochar for (10) minutes (Fig. 1). The statistical analysis's findings revealed that there were notable variations between weights and times (Table 1). The lowest removal rate was 55.09% at (10) minutes and a dose of (0.1) grams of biochar. The highest removal rate was 96.73% for cadmium at a concentration of 10 ppm in water treated with biochar at a dose of (0.5)gm and a time of (40) minutes (Fig. 2). The statistical analysis's findings revealed that there were no appreciable variations between weights and times (Table 2).



	Table 1:	Variance	analysis	test of	cadmium	(5)	ppm) treated	by biocha
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Tests of Between-Subjects Effects Dependent Variable: Cadmium										
Source	Type III Sum of Squares	Df	Mean Square	F	Sig.	Partial Eta Squared				
Corrected Model	1092.557ª	5	218.511	46.073	.000	.950				
Intercept	143837.449	1	143837.449	30327.965	.000	1.000				
Dose	692.168	1	692.168	145.943	.000	.924				
Time	301.214	2	150.607	31.755	.000	.841				
Dose * Time	99.175	2	49.587	10.455	.002	.635				
Error	56.913	12	4.743							
Total	144986.919	18								
Corrected Total	1149.470	17								
a. R Squared = .95	0 (Adjusted R So	uared =	= .930)							

 Table 2: Variance analysis test of cadmium (10 ppm) treated by biochar

Tests of Between-Subjects Effects										
Dependent Variable: Cadmium										
Source Type III Sum Df Mean Square F Sig. Partia										
	of Squares					Squared				
Corrected Model	4076.898ª	5	815.380	175.848	.000	.987				
Intercept	120195.411	1	120195.411	25921.750	.000	1.000				
Dose	2597.523	1	2597.523	560.191	.000	.979				
Time	1049.652	2	524.826	113.186	.000	.950				
Dose * Time	429.723	2	214.861	46.338	.000	.885				
Error	55.642	12	4.637							
Total	124327.951	18								
Corrected Total	4132.540	17								
a. R Squared $= .98$	7 (Adjusted R Sc	juare	d = .981)							

The lowest percentage of cadmium removal was (97.6%) at a dose of (0.1) gm of MBC and a time of (10) minutes. At a concentration of 5 ppm in water treated with MBC, the highest percentage of cadmium removal was (100%) at a dose of (0.5) gm of MBC and a time of (40) minutes (Fig. 3). The statistical analysis findings revealed that there were appreciable variations in weights and timings (Table 3). The lowest removal rate

was 90.36 percent at 10 minutes and 0.1 grams of MBC, and the highest removal rate was 99.96 percent for 10 ppm of cadmium in water treated with 0.5 grams of MBC and 40

minutes (Fig. 4). There were no appreciable variations between weights and times, according to the statistical analysis findings (Table 4).



Table 3: Variance analysis test of cadmium (5 ppm) treated by magnetic biochar(MBC).

Tests of Between-Subjects Effects									
Dependent Variable: Cadmium									
Source Type III Sum Df Mean F Sig. Partial Eta									
	of Squares		Square			Squared			
Corrected Model	12.039ª	5	2.408	6.540	.004	.732			
Intercept	177500.736	1	177500.736	482164.260	.000	1.000			
Dose	2.614	1	2.614	7.102	.021	.372			
Time	4.986	2	2.493	6.772	.011	.530			
Dose * Time	4.438	2	2.219	6.028	.015	.501			
Error	4.418	12	.368						
Total	177517.192	18							
Corrected Total	16.456	17							
a. R Squared = .732 (Adjusted R Squared = .620)									

Tabl <u>e</u>	4: Variance	analysis test	of cadmium ((10 pp	m) treated	by magnetic	biochar(MBC).
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Tests of Between-Subjects Effects										
Dependent Variable: Cadmium										
Source Type III Sum Df Mean Square F Sig. Partial										
	of Squares					Squared				
Corrected Model	255.375ª	5	51.075	113.955	.000	.979				
Intercept	168219.334	1	168219.334	375317.378	.000	1.000				
Dose	177.473	1	177.473	395.963	.000	.971				
Time	43.018	2	21.509	47.989	.000	.889				
Dose * Time	34.885	2	17.442	38.916	.000	.866				
Error	5.378	12	.448							
Total	168480.088	18								
Corrected Total	260.754	17								
a. R Squared = .979 (Adjusted R Squared = .971)										

The results of employing biochar to cadmium-contaminated remove aqueous solutions corroborated our findings by showing that biochar has the capacity to adsorb cadmium from aqueous solutions. (Yusuff et al., 2022; Jasim and Alzufri, 2022). The results showed a gradual increase in the removal ratio of cadmium at concentrations of 5 ppm and 10 ppm treated by biochar at a dose of 0.1 gm and 0.5 gm within 40 minutes at a constant temperature of 25-30 degrees Celsius. Due to the absorbent material's available surface area and the presence of active groups that aid in binding cadmium ions to it (Lei et al., 2019). While agglomeration of the micro-components of the absorbents, which reduces the distance between the micro-components of the absorbent materials and prevents the increase in efficiency, occurs at the lowest adsorption rate of cadmium at concentrations of 5 ppm and 10 ppm of cadmium ions when weighing (0.1) g of activated biochar at a time of (10)minutes (Kadhum and Albayati, 2022). It was also discovered that cadmium adsorption rates were higher with an increase in the adsorption dose of magnetic biochar compared to its predecessor biochar, where rapid uptake is related to the availability of active sites on the surface. These positive differences in results and adsorption efficiency were obtained and noted after adding Fe₃O₄ prepared in the laboratory to biochar to remove cadmium ions from the aqueous solution (Hassan et al., 2022), in addition to the primary removal mechanism, which is mostly accomplished by interactions functional with groups, electrostatic adsorption, and ion exchange between heavy metals and adsorbents (Luo et al., 2018). Figures 3 and 4 show that when we add magnetism to the biochar, the removal rates go up and the results are much better. This is because the sorbent material has a bigger surface area, its absorbing surface is better, and there are more places for the active grates to go. The highest removal ratio of cadmium

was recorded, and its concentration after treatment was 10 ppm (Wang et al., 2018). Finally, it was found that the adsorption process, whether for biochar or magnetic biochar for cadmium, decreases when the concentrations and weights used for treatment are increased. This is because the cadmium ion competes with the small area of the components of biochar and magnetic biochar for binding sites on the surface of the adsorbent material. While the lower concentration offers a driving force sufficient to overcome all resistances imparted to the solutes between the solid (biochar) and liquid (aqueous solution) phases, the adsorption resistance (Cd) lowers as the driving force for mass transfer increases (Aniagor et al., 2021). **FESEM and EDX Spectra Analysis:**

The biochar was examined after activation resulting from algae, and it was found that it is composed of basic elements such as carbon and oxygen, as shown by the FESEM image that appears in Figure (5). While Figure 6 shows a FESEM of the compound magnetic biochar, it also shows a picture of the elements from which the compound is composed: carbon, iron, and oxygen. Figure 7 is a field emission scanning electron microscopy (FESEM) analysis of the activated biochar (0.1 g) after treatment with cadmium ions (10 ppm) at a time contact of 40 min. The FESEM images show the morphology of the biochar surface after the activation process. On the other hand, FESEM images show the morphology of magnetic biochar (MBC) (0.1 g) after treatment with cadmium ions (10 ppm) at a time contact of 40 min (Fig. 8). Also, FESEM images could measure the nanoparticle size of the magnetic (Fe₃O₄), which is around 36 ± 6 . In addition, how these sit and homogenize distribution on the surface of biochar Moreover, we used Energy Dispersive X-ray analysis (EDX) as another technique to improve the composite structure of the BC and MBC.



Fig.5: FESEM of biochar activation before treatment.



Fig.6: FESEM of Magnetic biochar activation before treatment.



Fig.7: FESEM of cadmium ions (10 ppm) treated by 0.1 gm biochar after 40 minutes



Fig.8: FESEM of cadmium (10 ppm) treated by 0.1 gm magnetic biochar(MBC) after 40 minutes.

The EDX mapping and analysis are used to figure out the basic elements that

make up the compound. Furthermore, the EDX technique could give the percentage of

the element in the compound, so the users could determine the type of elements present and the percentage concentration of each element within the sample. Figure (9), is an EDX mapping and analysis of the elements of activated biochar (BC) (0.1 g) treated with cadmium ions at a concentration of 10 ppm, which shows the proportions of the different elements in the sample: cadmium (0.37%), carbon (90.56%), and oxygen (9.7%). While Figure (10), shows EDX mapping and analysis for the elements of magnetic biochar (MBC) (0.1 g) treated with cadmium ions at a concentration of (10) ppm, which shows the proportions of the different elements, cadmium (0.29%), carbon (82.89%), oxygen (15.3%), and Iron (1.53%),



Fig.9: EDX mapping and analysis of 0.1 gm biochar treated with Cd^{2+} at 10 ppm at 40 min.



Fig.10: EDX mapping and analysis of 0.1 gm MBC treated with Cd^{2+} at 10 ppm at 40 min.

XRD:

X-ray diffraction (XRD) is another technique to give more information about the structure of activated and magnetic biochar. Figure 11a is an XRD of activated and magnetic biochar. It shows peaks in the disordered carbon crystal plane of biochar activate (Cao *et al.*, 2012; Meng *et al.*, 2015). In addition, the peaks of iron oxide (magnetic nanoparticles) in magnetic biochar are at 30°, 35° , 43° , 53° , 57° , and 62.4° , respectively (JCPDS No. 33-0664) (Ruan *et al.*, 2015; Meng *et al.*, 2015). Figure 11b shows biochar activated and biochar activated after treatment with cadmium ions, which find extra low-intensity diffraction peaks at 54° and 73° of Cd-O (GCPDS No. 29-0713). Also, figure 11c shows magnetic biochar and magnetic biochar after treatment with cadmium ions to have extra low-intensity diffraction peaks at 54° and 73°, Cd-O (Yang *et al.*, 2018).



Fig.11: (a) XRD patterns BC and MBC, (b) $BC+Cd^{2+}$, (c) MBC+ Cd^{2+} at 10ppm with 0.1 g of biochar activate or magnetic biochar.

FTIR:

Fourier Transform Infrared spectroscopy (FTIR) is an important technique to determine the new function groups as a new structure in comparison with the structure before the reaction. Figure 12 is biochar before activation, which shows peaks at 3479 cm⁻¹, 3415 cm⁻¹ stretching, and 1066 cm⁻¹ for O-H bending in the water, 3238 cm⁻¹ for C-H stretching, 1622 cm⁻¹ for C=C stretching, and 1442 cm⁻¹ for C-C stretching in the aromatic ring. However, the biochar after pressing with hydrogen peroxide could activate functional groups such as hydroxyl and carboxyl on the surface. Thus, FTIR gives new peaks of 3370 cm⁻¹ for O-H stretching and 1386 cm⁻¹ for O-H bending of hydroxyl groups on the surface as phenol. Also, the broad peak from 3200–2600 cm⁻¹ for COOH stretching and 1055cm⁻¹ bending for C-O-H in carboxyl groups, figure 13, Moreover, the peaks at 1622 cm⁻¹ for C=C stretching and 1442 cm⁻¹ for C-C stretching in the aromatic ring have shifted to 1597 cm⁻¹ and 1412 cm⁻¹

respectively, because the conjugated groups of the aromatic ring overlap with new groups (Hurd, 1964; Sun et al., 2023). After the treatment with Cd²⁺ we found a reduction in the intensity of peaks 3371 cm⁻¹ and 3199 cm⁻ ¹, as well as a shifting of the bending peak from 1386 cm⁻¹ to 1367 cm⁻¹ for O-H of hydroxyl (phenol) because of the coordination bond with cadmium ions (Fig. 14). Not that all, the peak at 1055 cm^{-1} disappeared, and we see peaks at 2955 cm⁻¹ and 2922cm⁻¹ for coordination carboxyl groups with cadmium ions, which is attributed to the Cd-O of Cd²⁺ (Saha et al., 2007; Guo et al., 2019). Figure 15 is magnetic biochar, which shows a peak at 2958 cm⁻¹ and disappears from the bending peak at 1055 cm⁻ ¹ for coordination carboxyl groups with iron oxide (magnetite). The magnetite (Fe-O) stretching modes are represented by the absorption peak at approximately 561 cm⁻ ¹different peaks corresponding to different organic compounds were also identified (Hwang et al., 2014; Abderrahim et al., 2015;

Taha al.,2022; Jozanikohan et and Abarghooei, 2022). Furthermore, the peaks at 1597 cm⁻¹ and 1412 cm⁻¹ in biochar activate (Fig. 13) shifted to around the normal locations of 1605 cm⁻¹ and 1440 cm⁻¹ because it is not more conjugated with new functions groups (hydroxyl and carboxyl) after these are coordinated with magnetite. Because make coordination bond with cadmium ions is shown in Figure 14 we see peaks at 2953 cm⁻ and 2922 cm⁻¹ for coordination carboxyl 1 groups, which are attributed to the Cd-O of Cd^{2+} . Also, we found a reduction in the intensity of peaks 3371 cm⁻¹ and a shifting of the stretching peak to 3199 cm⁻¹ for O-H of hydroxyl (phenol) (Fig.16), because of the coordination bond with cadmium ions (Fourest *et al.*, 1994; Puranik *et al.*, 1999, Forsberg, 1988). According to earlier research, hydroxyl groups have a strong affinity for divalent cations. These hydroxyl and carboxyl groups, which are particularly abundant in biochar, can take on a negative charge and significantly aid in the adsorption of metals.



Fig. 12: FT-IR spectrum of biochar Spirulina platensis (BC)



Fig. 13: FTIR spectrum of biochar activate (BC activate)



Fig.14: FTIR spectrum of BC activate 0.5 gm treated with Cadmium ions at 10 ppm





Fig.16: FTIR spectrum of MBC 0.5 gm treated with Cadmium ions at 10 ppm

Conclusion

The adsorption of Cd^{2+} on biochar made from *Spirulina platensis* was shown to be dependent on a number of variables, including biomass dosage, contact time, and the initial concentration of Cd^{2+} . The best conditions for the algal biomass to remove cadmium ions were 0.5 g of biochar activate and MBC at 40 min of contact time, and initial cadmium concentrations of 10 ppm. The potential for removing cadmium ions from aqueous solutions with S. platensis biochar is enormous.

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